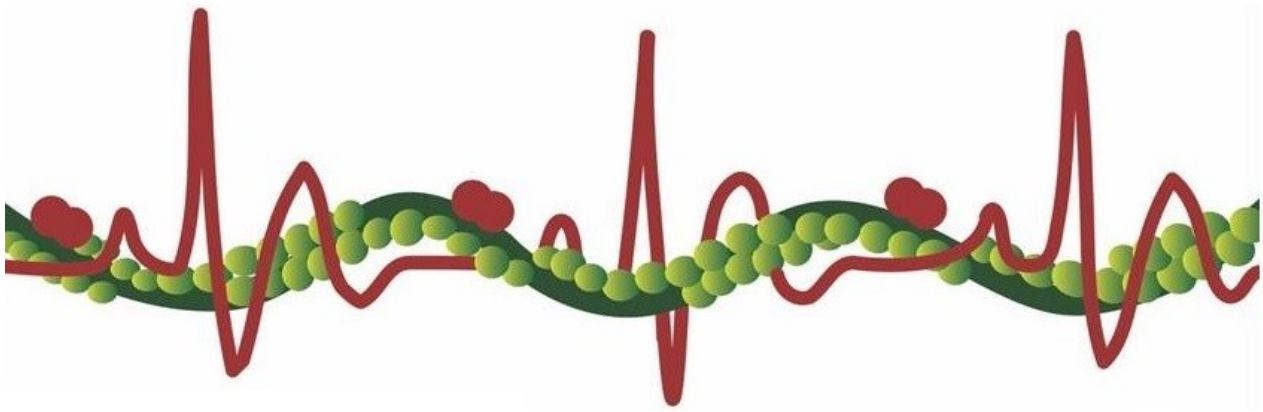


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UK National External Quality Assessment Service Cardiac Markers



Participants' Manual 2019



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1. General Information

1.1. Background

UK NEQAS Cardiac Markers is a “non for profit” centre working within NHS Greater Glasgow and Clyde to improve cardiac biomarker investigations in the clinical laboratory sector for the benefit of patients. Greater Glasgow and Clyde established under the National Health Service (Scotland) Act 1978, known as NHS Greater Glasgow & Clyde (Legal Entity), operating UK NEQAS Cardiac Markers is a UKAS accredited proficiency testing provider (NO:8560) (BS EN ISO 17043:2010).

UK NEQAS Cardiac Markers EQA Scheme comes under the organisational framework of UK NEQAS and follows the UK NEQAS code of practice for member schemes, but is financially and organisationally distinct. UK NEQAS Cardiac Markers is supported financially by fees paid by participants. This allows the scheme to run independently.

1.2. Scheme Management

The schemes for cardiac markers are run from the Laboratory Medicine & FM Building, Queen Elizabeth University Hospitals, situated within the Biochemistry Department. It is managed by Alan Reid (Scheme Organiser), who will be pleased to provide advice or assistance on any aspect of the scheme or visit our web site at www.ukneqas-cm.org.uk

1.3. ContactDetails

UK NEQAS Cardiac Markers
Level 1 (Room B/046)
Laboratory Medicine & FM Building
Queen Elizabeth University Hospital
1345 Govan Road, Glasgow
G51 4TF Scotland UK

General e-mail: info@ukneqas-cm.org.uk

Website: www.ukneqas-cm.org.uk

Phone: +44(0)141 440 2888

Fax: +44(0)141 440 1274

Alan Reid (Scheme Organiser)

e-mail : alanreid@ukneqas-org.uk

Maggie Forsyth (Quality Manager)

e-mail: maggie@ukneqas-cm.org.uk

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1.4. Objectives

UK NEQAS Cardiac Markers aims to:

- Provide an accurate, fair, efficient, cost effective UKAS accredited external quality assurance programme to all participants of the scheme within the resources available, therefore assuring a high quality diagnostic service to the participants' patients
- Provide professionally-led and scientifically based EQA scheme with a primarily educational objective
- Provide a regular, reliable distribution of appropriate specimens
- Provide rapid feedback of individual participant performance in reports that are comprehensive and readily understood
- Provide Data on method related performance
- Facilitate laboratories of participating members to fulfill ISO 15189:2012 requirements regarding participation in inter-laboratory comparison programmes.
- Maintain and improve performance of diagnostic testing at a high level of proficiency, wherever testing is performed.
- Provide an opportunity to participate in EQA as an established part of Quality Assurance, actively encouraged by professional bodies.

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1.5. Design overview

UK NEQAS for Cardiac Markers is organised to provide an External Quality Assessment (EQA) Scheme for the assay of serum cardiac Troponin I and Troponin T, Creatine Kinase (CKMB), Myoglobin, B-type Natriuretic Peptide (BNP) and N Terminal -ProB-type Natriuretic Peptide (NT-proBNP) and will develop appropriate EQA schemes for new cardiac biomarkers.

The fundamental design of the scheme began in 1997 there the requirement for a UK wide EQA scheme for the new cardiac markers Troponin I (cTnI) and Troponin T (cTnT) was recognised and a scheme for these two analytes was proposed. This received recognition and funding as a pilot scheme from Clinical Pathology Accreditation (UK) Ltd (CPA) in February 1998.

The cardiac Troponin EQA schemes received CPA accreditation in December 1999. In June 2002 Myoglobin and CKMB Point of Care Schemes also received CPA accreditation. These developments were supported from participant fees. We have developed an EQA material suitable for N terminal Pro-B type Natriuretic Peptide (NT-ProBNP) and B-Type Natriuretic Peptide (BNP) on both laboratory based systems and Point of Care systems. On the 23rd March 2016 the scheme became United Kingdom Accreditation Service (UKAS) accredited to ISO 17043 General Requirements for Proficiency Testing.

All of these schemes are now established in the UK and our priority is to continually maintain their quality and where appropriate develop EQA schemes for new cardiac biomarkers. Maintaining accreditation to ISO 17043:2010 is seen as an essential step in meeting these aims.

UK NEQAS Cardiac Markers is a simultaneous participation proficiency testing scheme, distributing to participants for concurrent testing. Within UK NEQAS Cardiac Markers the frequency of distribution is 12 per year as documented in the Distribution Schedule. A 'distribution' basis may be two, three or four specimens' dependant on the specific Scheme. After completion of the testing, the results are returned to UK NEQAS Cardiac Markers and compared to give an indication of the performance of the individual participants to the group as a whole.

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1.6. Schemes

Cardiac Marker Schemes (Laboratory Based Scheme)	
Number of samples (All Methods, all analytes):	Three lyophilised sample per distribution.
Additional Sample for high sensitivity Methods (Troponin):	One lyophilised sample per distribution
Frequency of distribution:	Twelve per year
Range of tests:	Cardiac Troponin I, Cardiac Troponin T, CKMB, Myoglobin, and NT-pro <i>B-type Natriuretic Peptide</i> (NT-proBNP).
Concentration ranges:	Varied by analyte additions
Troponin I and T additions are in the form of a complex. This complex is released in to the bloodstream of patients with myocardial muscle cell injury and have the required source specificity. CKMB and Myoglobin also have required source specificity. NT-ProBNP is a recombinant peptide which has the required source specificity.	
Concentration ranges employed in this EQA Scheme(cTnI, cTnT, CKMB and Myoglobin)	
These cover the clinically encountered range from less than the assay detection limit, levels around the diagnostic decision limits and levels found in the 24 hours following a myocardial infarction or after thrombolysis wash out. For high sensitivity Troponin methods 4 th sample will be in the range of 3 – 20 ng/L. The numerical value depends on the assay method.	
Concentration employed in this EQA (NT-proBNP)	
These cover the clinically encountered ranges from less than the assay detection limit, levels around the diagnostic decision limits and levels found in patients with heart failure For NT-proBNP the numerical value depends on the assay method.	
Base Matrix:	Pooled human serum (female only).

B-Type Natriuretic Peptide (BNP) Scheme (Laboratory Based Scheme)	
Number of samples :	Three Lyophilised samples per distribution.
Frequency of distribution:	Twelve per year
Range of tests:	<i>B-type Natriuretic Peptide</i> (BNP)
Concentration ranges:	Varied by analyte additions
BNP additions are in the form of a synthetic peptide which has the required source specificity.	
Concentration employed in this EQA (BNP):	
These cover the clinically encountered ranges from less than the assay detection limit, levels around the diagnostic decision limits and levels found in patients with heart failure For BNP the numerical value depends on the assay method.	
Base Matrix:	Pooled human EDTA plasma (female only).
Diluent:	One vial containing 2 mL diluent supplied with each distribution

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Point of Care Scheme (Serum) (CMKPS)	
Number of samples (All Methods, all analytes)	Two lyophilised samples per distribution + 500 µL of diluents supplied in plastic vials for each sample.
Frequency of distribution:	Twelve per year
Range of tests:	The EQA covers cardiac Troponin I, cardiac Troponin T, CKMB, Myoglobin and NT-pro <i>B-type natriuretic peptide</i> (NT-proBNP)
Concentration ranges:	Varied by analyte additions
Troponin I and T additions are in the form of a complex. This complex is released in to the bloodstream of patients with myocardial muscle cell injury and has the required source specificity. CKMB and Myoglobin also have required source specificity NT-ProBNP is a recombinant peptide which has the required source specificity.	
Concentration ranges employed in this EQA Scheme(cTnI, cTnT, CKMB and Myoglobin)	
These cover the clinically encountered range from less than the assay detection limit, levels around the diagnostic decision limits and levels found in the 24 hours following myocardial infarction or after thrombolysis wash out. The numerical values depend on the assay method.	
Concentration employed in this EQA (NT-proBNP)	
These cover the clinically encountered ranges from less than the assay detection limit, levels around the diagnostic cut-off and levels found in patients with heart failure. For NT-proBNP the numerical value depends on the assay method.	
Base Matrix:	Pooled human Serum (Female only)

Point of Care Scheme (Plasma) (CMKPE)	
Number of samples (All Methods, all analytes)	Two lyophilised samples per distribution + 500 µL of diluents supplied in plastic vials for each sample.
Frequency of distribution:	Twelve per year
Range of tests:	Cardiac Troponin I, CKMB, Myoglobin, and <i>B-type Natriuretic Peptide</i> (BNP)
Concentration ranges:	Varied by analyte additions
Troponin I additions are in the form of a complex. This complex is released in to the bloodstream of patients with myocardial muscle cell injury and have the required source specificity. CKMB and Myoglobin also have required source specificity. BNP is a synthetic peptide which has the required source specificity.	
Concentration ranges employed in this EQA Scheme(cTnI, CKMB and Myoglobin)	
These cover the clinically encountered range from less than the assay detection limit, levels around the diagnostic decision limits and levels found in the 24 hours following myocardial infarction or after thrombolysis wash out The numerical values depend on the assay method.	
Concentration employed in this EQA (BNP)	
These cover the clinically encountered ranges from less than the assay detection limit, levels around the diagnostic decision limits and levels found in patients with heart failure. For BNP the numerical value is depends on the assay method.	
Base Matrix:	Pooled human EDTA Plasma (Female only)

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1.7. The number and type of expected participants

Laboratory Based Schemes (August 2019)

Analyte	Number of Participants
Cardiac Troponin I (inc "hs" cTnI)	223
Cardiac "hs" Troponin T	240
CKMB (Mass)	29
CKMB (Activity)	6
Myoglobin	25
BNP	48
NT-proBNP	162

Point of Care Based Schemes (August 2019)

Analyte	Number of Participants
Cardiac Troponin I	25
Cardiac Troponin T(Roche Systems)	9
CKMB(Alere Triage)	13
Myoglobin(Alere Triage)	13
BNP	14
NT-proBNP	13

1.8. Changes to Schemes

UK NEQAS Cardiac Markers shall promptly advise participants of any changes to EQA Scheme design or operation. Various aspects of the EQA Programme can from time to time be subcontracted. UK NEQAS Cardiac Markers does not subcontract the planning of the EQA Programme, the evaluation of performance or the authorisation of the final report. If any subcontracting is necessary, UK NEQAS Cardiac Markers will be responsible for ensuring the quality of this work.

1.9. Participant Appraisal of the Service

The scheme organises educational and participant meetings as well as annual questionnaires to allow participant appraisal of the service. This aids to inform decision making and to set quality objectives to improve the scheme.

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1.10. External Regulation of Our Services

Accreditation

UK NEQAS Cardiac Markers is accredited by UKAS against the requirements of ISO/IEC 17043:2010 – Conformity Assessment – General requirements for proficiency testing. The scope of accreditation can also be viewed on the UKAS website: [Scope of Accreditation](#)

Steering Committees & Specialist Advisory Groups

All EQA providers are required to seek advice from and report to Steering Committees and/or Specialist Advisory Groups. For Terms of Reference see Appendix 1. The Clinical Chemistry division of UK NEQAS is presently served by an overall Steering Committee (SC) which advises on overall policy matters, with Specialist Advisory Groups (SAGs) providing external scientific advice.

Steering Committees do not consider the performance of individual participating laboratories, except in advising on performance criteria or where this may indicate a failure in the operation of the Scheme (and even in such cases the laboratories will not be identifiable). For current membership of Steering Committee see Appendix 2

The Steering Committee (Clinical Chemistry) meets twice a year, and the Specialist Advisory Groups: Immunoassay three times a year. For current membership of Specialist Advisory Groups: see Appendix 3

National Quality Assurance Advisory Panel

The Panels are independent bodies of representatives nominated by the professions whose role is to monitor standards of laboratory performance in the UK. The UK NEQAS Cardiac Markers Scheme Organiser will co-operate with the Panel as required (see Appendix 4). A member of the Panel is represented on the Steering Committee and the Specialist Advisory Group.

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2. Terms and Conditions of Participation

2.1. Eligibility

The scheme provides services to all clinical and research laboratories and point of care providers in the UK and abroad. Manufacturers may do so 'anonymously', whilst a product is not yet available on the market, or on an 'information only' basis, i.e. without receiving samples and returning results.

Intending participants can access registration information on the UK NEQAS Cardiac Markers [website](#) or can contact the Scheme to request information.

2.2. Terms and Conditions

The act of requesting registration or re-registering confirms a participant's willingness to be bound by these terms and conditions. For UK clinical laboratories, the act of enrolling in a scheme confirms their willingness to be bound additionally by the current [Joint Working Group for Quality Assurance](#) Conditions of Participation.

Any manufacturer whose assay is represented in the EQA programmes also agrees to be bound by the JWG Conditions of Participation and may be brought to the attention of the Medicines and Healthcare products Regulatory Agency (MHRA) in the event of poor assay performance

- EQA samples must be treated in an identical manner to a laboratory's routine clinical samples
- Participants must inform UK NEQAS Cardiac Markers of any method changes
- Failure to pay subscription fees on presentation of an invoice may result in discontinuation of participation and referral to the relevant NQAAP
- Collusion between participants and falsification of results is not permitted.

2.3. Period

Participation in all UK NEQAS Cardiac Markers Schemes is deemed to be continuous with automatic annual renewal and invoicing for subscription fees for each NHS financial year (1st April to 31st March). Participation may begin at any time during the year; part-year charges are higher than pro rata.

2.4. Participation fee

We recognise the financial constraints being imposed upon many laboratories and therefore we keep our participation fees as low as possible.

The participation fee for the Laboratory Based Schemes and Point of Care Schemes are available on request. This fee covers all sample distributions and associated reports for a 12 month period, starting on 1st April. All postage costs are included within the subscription fee. Overseas participants' fees only cover air-mail postage and additional charges may be required for courier delivery. VAT charges will be made appropriately as required by HM Customs and Excise at 20%. Once a purchase order has been received an invoice will be issued by our host organisation and Legal entity, NHS Greater Glasgow & Clyde.

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A participant may withdraw from the Scheme at any time, but no refund will be given of fees paid.

If further information is required about the fee structure please phone: +44 (0)141 440 2888 or email: info@ukneqas-cm.org.uk

2.5. Confidentiality

Information provided by participants is treated as strictly confidential as outlined by UK NEQAS Cardiac Markers Privacy Policy. Participant identification will be known only to persons involved in the operation of the Scheme. Participant will not be released to third parties, unless written approval has been provided to waive confidentiality.

Participant details and performance data are only revealed as part of the poor performer's procedure for the UK and the Republic of Ireland. In this situation there is a formal procedure to address the issues, the final step of which could result in the participating laboratory being referred to the National Quality Assurance Advisory Panel. Laboratories providing a clinical service in the UK are governed by the conditions of confidentiality determined by the Joint Working Group on Quality Assurance (Appendix 1).

UK NEQAS Cardiac Markers reports are copyright and may not be copied, distributed, published or used for publicity and promotion in any form without the written consent of the Scheme Organiser on each and every occasion, though performance data may be shared with individual clients (eg GPs, clinicians, pharmaceutical companies) without consultation.

2.6. Repeat Samples

Limited numbers of single samples or sets from a particular distribution are usually available to full participants who may wish to check aberrant results from previous distributions or require fresh samples from current distribution due to sampling errors. UK NEQAS Cardiac Markers reserves the right to ask why repeat samples are needed and limit their supply if this would compromise the service to other participants

2.7. Reporting of Results

All full participants are expected to return results promptly within the specified reporting period. Those under the remit of the UK NQAAP are expected to return 100% of results within the relevant cumulative performance scoring period. Where a laboratory is unable to return a set of results, an explanation must be provided using the comments box on the online result entry page.

2.8. Demographic information

Participants are responsible for ensuring that the contact (including invoicing contacts), method and reporting units information held by UK NEQAS Cardiac Markers is current and up-to-date. Method and unit information can be communicated via phone: +44 (0)141 440 2888 or email: info@ukneqas-cm.org.uk

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3. Materials

UK NEQAS Cardiac Markers use lyophilised samples which are prepared in house, including homogeneity and stability testing. All suppliers are evaluated for their competency to provide these services. All samples should be treated as if they were patient samples. All preparation is carried out to ISO17043 specifications that ensure uniformity and condition of samples. EQA samples should be regarded as possibly infectious material and handled according to local procedures.

3.1. Pool Manufacture

Pools are prepared using a pooled female human plasma or serum base matrix. Analyte levels are varied by addition of Troponin complex, CKMB, Myoglobin, NT-pro-BNP and BNP to the base matrix. All materials have been screened for infective agents. Samples, included within a distribution, are lyophilised and have a reconstituted volume of 500 µL.

3.2. Participant Handling and Storage of EQA Materials

Lyophilised samples should be reconstituted by addition 500 µL of distilled or deionised water using a pipette with a tolerance of ±8 µL. Additional plastic vials with pre-measured diluent are supplied with the BNP scheme. Vials should be allowed to stand for 15 minutes with occasional mixing and then stored at 4°C.

The EQA material should be assayed after reconstitution as follows:

- cTnI, cTnT, CKMB, Myoglobin and NT-proBNP between 1 and 3 hours.
- BNP between 40 and 90 minutes post reconstitution.

For the Point of Care based scheme 2 plastic vials containing appropriate volumes of diluent are provided for reconstitution in locations out with the laboratory. Instructions on reconstitution of the materials are given on the report form enclosed with the distribution. Samples should be treated in the same manner as clinical samples.

3.3. Stability

It has been established that at 4°C, troponin (I and T) loss is <2% over 3 hours. CKMB, Myoglobin and NT-proBNP are also stable for this period of time. However BNP is a labile peptide and should be assayed between 40 –90 minutes post reconstitution.

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4. Operations

4.1. Distribution Cycle

All schemes operate according to a regular cycle of activity with 12 distributions per year. A distribution has a unique identifier (usually numeric) with fixed sample despatch and results return dates.

The distribution schedule is available on the [website](#) (and at participants' request) from the beginning of each year. This schedule indicates the date that samples are issued, the closing date for return of results and the date that reports are issued to give sufficient notice of Scheme activities. Dates may be subject to minor changes dependent upon operational circumstances.

Participants should contact the Scheme if they have not received their monthly distribution (all or part), or their samples have been damaged in transit. UK NEQAS Cardiac Markers will replace the non-received EQA material(s) and initiate an investigation of the non-Conformance to avoid repeat occurrence.

4.2. Method Classification

A crucial element of participation for all schemes is the correct assignment of method codes, since performance scoring may be method-based, and the provision of accurate method-related information is an important element of the service. Methods are given unique codes in the computer system. Considerable effort has to be expended by UK NEQAS Cardiac Markers staff to ensure the accuracy of method coding and updating records when these change. Participants are required to cooperate with this process by informing us of errors, omissions or changes at the earliest opportunity.

4.3. Packaging & Mailing

Samples for each distribution are mailed to the registered scheme contact(s) using the fastest and most appropriate route possible. First class mail is used for the UK, and airmail (with express surcharge where necessary). The packages are clearly marked 'EQA Samples URGENT'. Packaging complies with current UK legislation for the mailing of pathological material.

All tubes are labelled with the scheme, distribution identifier and sample number. The naming convention for the latter comprises a sequential numeric distribution identifier plus a letter where there are multiple specimens in a distribution (e.g. 256A, 256B, 256C). Once the specimens have been despatched, the registered scheme contacts are sent an e-mail notification of sample despatch.

If for any reason specimens are not received, not received when expected, or are damaged upon receipt, notify UK NEQAS Cardiac Markers via one of the methods listed under 1.3 Contact Details.

4.4. Results Documents

All schemes have distribution-specific Report Forms which are individual to each participant. These carry the laboratory code, a full list of the analytes available in the scheme, as well as details about sample handling and return of results. In some cases method confirmation is also included. They are under constant review to make them easy to understand and use, and may change from time to time to reflect improvements. The functionality of the Report Form mirrors the Results return section of the online service.

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4.5. Results Reporting Procedure

Result entry, via the WES result entry/ reporting portal ([WES result entry / reporting portal](#)), opens on the distribution date and the result sheet distributed with samples details this. Participants are notified by e-mail when the distribution opens. Each UK NEQAS number is assigned a unique password to access the WES system, which is provided upon commencement of participation along with a username and password for the UK NEQAS Cardiac Markers website.

Unless specified otherwise, all results must be returned using our online service. This provides the most rapid means of reporting results, with real-time validation against plausibility limits and confirmation of submission.

In the unlikely event that the online results entry system is not available, results transcribed onto the original Report Form document may be FAXED or sent as an e-mail attachment, **by prior arrangement with UK NEQAS Cardiac Markers**. Participants should give date of receipt of samples on submission of results.

Results should be entered in the units shown on the online service form, taking care to match sample numbers and avoid transcription or transposition errors. If the units displayed on your results document are different to your usual reporting units, please notify us.

- **Late results / returns** are defined as results received after the distribution has closed, but before the report is published. Participants should return late results along with an explanation of why results were returned late. Late returns are monitored by UK NEQAS Cardiac Markers.
- **Null returns** where there are no results available e.g. if the assay is out of service, or if one or more results from a panel of analytes is missing, then enter XPL into the results entry boxes for the affected analytes/specimens and provide a brief explanation in the comments box at the bottom of the results entry screen.

- **Failure to return results**

When a participant fails to return results for 3 distributions within the 12 month scoring window they will be contacted by the Scheme Organiser and may be referred to the appropriate NQAAP as a Persistent Poor Performer.

4.6. Amendments Prior to Data Processing

Participants who discover an error in their reported results before the reporting deadline can amend their results via the online service at any time whilst the distribution is open.

4.7. Amendments After Data Processing

If errors are identified after the return-by date, requests to amend non-analytical errors should be made by contacting the Scheme in writing. Requests for amendments can only be made for the distribution prior to the one currently open. The requestor should include their name and a valid reason

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- **Late results / returns** are defined as results received after the distribution has closed, but before the report is published. Participants should return late results along with an explanation of why results were returned late.
- **Very Late Returns** are defined as results received after the distribution has closed, and after the report is published. Very Late Returns will only be accepted if they are accompanied by evidence of results (e.g. a lab worksheet or computer printout) and an explanation of why results were returned very late.
- **Blunders** - Participants who have submitted an incorrect result due to a non-analytical error (e.g. a transcription error when entering results online, or a unitage error) may contact UK NEQAS Cardiac Markers to have their results and report amended. They must state the Scheme, Distribution number and reason(s) for the error, and copies of results obtained (lab worksheet or computer printout) must be appended.

Participants will be investigated:

- if they submit "very late" results
- if the action for a blunder is a request to amend results / reports without appropriate evidence supplied
- on the basis of reliable reports of suspected collusion / falsification of results.

If the suspicion of collusion / falsification of results is supported by strong evidence, the participant will be suspended and all reports will be halted until further notice. If collusion is proven, the relevant competent authority will be notified.

Amending results is at the discretion of the Scheme Organiser and is not an automatic entitlement. The Organiser reserves the right to request additional evidence in the form of the original output from the analyser. All amended results are flagged as such in our database and monitored.

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5. Data Processing

5.1. Data Handling

All Scheme data are held on secure network servers which are backed up daily. Data processing is performed using bespoke EQA software modules which have been developed in association with the Wolfson Computer Laboratory.

5.2. Calculation of Target Values

As there is no accuracy base and no internationally recognised reference preparations currently exist for the analytes in this scheme consensus values are used. Validity of these consensus means will be assessed by reissue of pools over different time periods.

To eliminate the distorting effect of grossly atypical results, outliers are trimmed from both tails of the ranked data set, with a corrected estimate of dispersion (SD or CV) by the method of Healy (1979)² to allow for the removal of extreme values which are not 'true' outliers. The data processing for individual schemes is conducted using individually configured modules within the computer system.

For all our Schemes we have a minimum data set size that we use to calculate a target value. We are aware that for analytes that have a low number of users, those participants do want to see some level of assessment even though both they and UK NEQAS Cardiac Markers are well aware of the limitations of this approach. Statistically speaking, this carries a huge uncertainty overhead but the alternative is not to provide Laboratories with any indication of their performance.

For those analytes with an ALTM target that regularly Comprises Fewer Than 10 Results, There Is Often No Formal Performance Assessment. See Section 6.8 For Minimum Participant Numbers.

5.3. Calculation of Performance Scores

As well as providing data on closeness to the target value in a given distribution, schemes employ scoring systems which yield a performance score averaged over a number of distributions and individual samples within a rolling time window to give a robust estimate of overall bias and its variability.

The scoring method used for all UK NEQAS Cardiac Markers schemes is the 'ABC of EQA' system. Unlike many other schemes which conduct a series of discrete cycles after which an 'end-of-cycle report is prepared, UK NEQASs operate on the basis of continuous analysis for a rolling time window.

5.4. Acceptable Performance Criteria

Limits for acceptable performance scores are recommended by the Scheme Organiser and endorsed by the NQAAP after due deliberation and consultation with Organiser and SAG, to reflect the state of the art of analysis and encourage improvement. Procedures are used to identify those laboratories which have breached these limits on a set number of occasions within the cumulative reporting period. Schemes are required to provide information on persistent poor performers to the National Quality Assurance Advisory Panel (NQAAP) for Chemical Pathology. See Section 8 Complaints and Appeals Procedure

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5.5. Processing Surveillance

As each distribution is processed, the Scheme Organiser carefully checks the resulting data for integrity and consistency of results, and any unexpected shifts in method-related values which might signal a clinically-important shift in diagnostic products.

6. Reports and Report Interpretation

6.1. Target Turn-Around Times for Reporting

Reports are issued to participants in accordance with the Distribution Schedule (available on the UK NEQAS Cardiac Markers [website](#)). UK NEQAS Cardiac Markers target for report issue is within 1 week of distribution closure. The exact time is recorded and is regularly audited.

6.2. Report Versions

The current definitive report is that on the secure Results and Reports website. When reports for a scheme distribution are published the name and date of authorisation are stated, and the time and date of publication are printed on each page. Unless specified in the report (e.g. an initial report awaiting republishing with a commentary) this may be considered as a 'final' report.

Revised reports including a participant's late or amended results are identified by the words "LATE RESULTS" or "AMENDED" below the laboratory code at the top right of the report. Changes in target values resulting from inclusion of late or amended results are normally trivial (and will always improve target validity), and will be reflected in reports for subsequent distributions. On the rare occasions when revised reports are published for a distribution, the revised report is clearly identified as such, with reference to the date when the report which is replaced was published.

If additional data is received and flagged as a late return:

- If the method group has less than 10 participants an amended report will be issued to all participants using this method with a comment as to why they have received this report and indicating that this supersedes the previous report. Date of previous report will be given.
- If the method group has greater than 10 participants the Scheme Organiser or deputy will assess whether there has been a significant change in method target which will affect resultant statistics. If so then amended reports will be issued, as described above, to all participants using the affected method. If not, an amended report will be issued to the late returning laboratory.

6.3. Distribution of Reports

Reports can be accessed by participants by logging in to the WES result entry/ reporting portal ([WES result entry / reporting portal](#)). Participants will be notified by e-mail when reports are available. Each UK NEQAS number is assigned a unique password to access the WES system, which is provided upon commencement of participation along with a username and password for the UK NEQAS Cardiac Markers website. In the unlikely event that reports are not issued in accordance with the Distribution Schedule, participants will be notified.

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6.4. Report Formats

Schemes' reports are the main interface with participants, and a great deal of effort has gone into making these informative and easy to interpret. All scheme reports are generated as A4 format PDF files, which display the data in a number of discrete tabular and graphic formats shared across related schemes.

Scheme reports now have 'traffic light' colour coding, where symbols and their colour (green, yellow or red) indicate how close individual percentage biases are to the target value, and whether performance scores lie within or outside acceptable limits. Examples are available on request.

- **Reports consist of the following pages**

- ... Feedback
- ... Performance Summary Icons
- ... Participation summary
- ... Distribution Summary
- ... Method Summary
- ... Cumulative Summary
- ... Cardiac Troponin I
- ... hs Troponin I (4th sample)

In the above example there are reports for Cardiac Troponin I and hs Troponin I (4th Sample). These method pages are associated with analytes participant's are registered for on the scheme.

- **Feedback Page**

Address and Comments

Any comments you made to us are shown on this page and have been acted upon where necessary

Any specific comments applicable only to laboratory XXXXX are shown on this page

Any general comments applicable to all laboratories are shown on this page

General comment: Report authorised by Alan Reid (Scheme Organiser)

6.5. Result Validation

UK NEQAS Cardiac Markers will send an email notification to the registered scheme contacts when pdf reports for a scheme have been published to the website. Each report is specific for the laboratory identifier and password entered.

The results for that distribution should be checked by participants to ensure that they are the ones returned by your laboratory.

Mistakes can occur. Requests to amend non-analytical errors should be made directly contacting UK NEQAS Cardiac Markers. Amending results is at the discretion of the Scheme Organiser and is not an automatic entitlement. The Organiser reserves the right to request additional evidence in the form of the original output from the analyser. All amended results are flagged as such in our database and monitored.

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Also, it is crucially important that participants' methods (and sub-methods, instruments, reagents, calibrants and reporting units where appropriate) are accurately identified, especially where performance is assessed against the method mean. Any apparent discrepancies should be corrected by contacting UK NEQAS Cardiac Markers directly.

6.6. Definitions

The various indices used in the scoring system may be defined as:

All Laboratory Trimmed Mean (ALTM): The recalculated mean value after exclusion of all results outside 2 (or 3) SD from the All Laboratory Mean.

- Troponin T
- Myoglobin
- CKMB

Method Laboratory Trimmed Mean (MLTM): The recalculated mean value of results returned by all laboratories using the same method. Widely discrepant results are trimmed as for the ALTM.

- **Troponin I & BNP**(methods the lack of agreed standardisation increases method to method variability beyond that usually encountered in established immunoassay i.e. that arising from variations in antibody - epitope responses. The calculation of an all method trimmed mean (ALTM) is calculated for information but participants are scored against their MLTM.

Group Laboratory Trimmed Mean (GLTM): As for MLTM but using all results from laboratories with related methods which have been predefined into a method group. Data will not be trimmed if <20 data points.

- **NT-proBNP**

6.7. ABC of EQA Scoring

'ABC of EQA' is an ISO Guide 17043 compliant framework which meets and surpasses the utility of existing systems. The main benefit for participant laboratories, EQA Organizers, Steering Committees, Specialist Advisory Groups and the NQA Advisory Panels alike, is that it is a single system, which not only works across analytes, schemes and disciplines, but can allow meaningful comparisons to be made between analytes, schemes and disciplines.

External quality assessment (EQA) is intended to give you an independent and objective assessment of your performance. This requires effective scheme design, including

- a reliable basis for assessment, with reliable specimens and valid target values.
- sufficient recent data, achieved by frequent distributions and rapid turnaround of reports
- effective communication through informative, intelligible reports and a running scoring system

UK NEQAS reports are structured so as to complement and best utilise the 'ABC of EQA' scoring system, and reports in the UK NEQAS 'house style' allow you to see at a glance if you are performing well. If you are performing well, no further action is required. If you are not performing well then you can probe

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further into the data presented. Similarly, you can see if you are performing in keeping with other users of your method and whether the method itself is performing well.

There are three scores **A, B and C**

A is for Accuracy (total error)

B is for Bias

C is for Consistency of bias

which are conveniently referred to as the '**A score**', '**B score**' and '**C score**', or simply **A, B and C**.

- Every laboratory will have an **A, B and C** score for **each analyte** they measure.
- **All 3** scores should be used when assessing performance.
- The **B** and **C** scores (which have not been transformed) are best looked at together and provide analytical data on average bias and its consistency (pattern).
- The **A** score is weighted as part of a transformation process to take into account factors such as 'degree of difficulty' and normalised (median set at 100)
- The **A** score is primarily used as a quick 'comparator' or 'screening tool' for performance across all analytes. An **A** score of '100' is 'average', but this may of course be 'better' or 'worse' than what is required clinically, depending on the analyte.
- As more UK NEQAS schemes adopt the '**ABC of EQA**' approach, the more useful the **A** score becomes in allowing broad comparisons to be made between analytes

6.7.1. A, B and C scores in detail

Each of the 3 scores is calculated over a **rolling time-window** and thus comprises data (results) from many specimens. They are always being updated with fresh current data, while older data drops out of the 'time-window'. The time-window has been set at 6 distributions (equivalent to 6 months) for 'standard' schemes.

For **all** UK NEQAS Cardiac Marker Schemes and in line with other UK NEQAS centres, all scores are set so that **a low score is 'good', a high score is 'bad'**.

- The Accuracy **A score** tells you, on average, how good your overall performance is. This takes into account such factors as **bias, consistency of bias, degree of difficulty** etc. It has been transformed to ensure that **A scores** are broadly equivalent across analytes. For example, if you have an **A score** of 85 for Troponin I and you also have an **A score** of 85 for BNP, this would indicate that you are performing both, on average, equally well.
- The Bias **B score** tells you how far away from the 'target', on average, you are. It has not been transformed.
- The Consistency of bias **C score** tells you, on average, if you usually have the same bias pattern. It is also not transformed and can assist in answering the following questions. '*Do you have different bias depending on the concentration of analyte in the sample?*' '*Does your bias vary depending on the specimen matrix?*' '*Has your bias changed during the time window?*' '*Are you imprecise?*' A high (poor) **C score** does not necessarily mean that you are imprecise, though if you **are** imprecise, it is impossible for you to have a very good (low) C score. *Poor consistency of bias is **not** the same as imprecision.*

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6.7.2. A, B and C score calculation

The specimen-level % bias calculation (**specimen %bias**) is at the heart of all calculations:

$$\text{specimen \% bias} = \frac{(\text{result}-\text{target})}{\text{target}} \times 100$$

If the target is 10 and you get a result of 11, then your bias is +10%; if the target is 10 and you get a result of 8, then your bias is -20%; if the target is 10 and you get a result of 10, then your bias is 0%, and so on.

We then calculate your '**B score**', (i.e. your average bias), as the **trimmed mean** of all individual '**specimen %biases**' (including the sign) in the rolling time window.

The '**C score**' is simply the **standard deviation** (adjusted to take into account the degree of trimming) of the data which make up the B score.

The **A score** is an estimate of **accuracy [total error]** in UK NEQAS and is derived as follows:

- we take your **Specimen % bias** and transform it by a 'degree of difficulty' factor (see below) to get your **Specimen transformed bias** [this can be positive or negative]
- we then take the modulus of this **Specimen transformed bias** to give the **Specimen Accuracy Index** [as it is a modulus it has no sign]
- finally, we calculate your '**A score**' as the **trimmed mean** of all of your **Specimen Accuracy Indices** in the rolling time-window.

6.7.3. How did we choose the 'degree of difficulty factor'?

The transformation itself has been empirically derived separately for each analyte and is based on modelling of data dependent on the concentration (target value) for the individual specimen. We did this by:

- examination of **the relationship between CV and target value** for the analyte, based on 2008-2009 data
- **derivation of an equation** for this relationship, to yield concentration-dependent factors
- **normalisation** of the factors to yield a median A score of 100 at January 2010

6.8. Minimum Participant Numbers

See section 5.2 Calculation of Target Values

The minimum number of participants required to produce meaningful statistics is >10 for all analytes.

Reports will reflect this by including this statement:

"The scoring of a method or a group of methods with < 10 participants should be considered as out-with scope of our current accreditation." Also reference that a participant should refer to the participant manual.

This statement will appear on the following pages of the report:

- Distribution Summary Page
- Each analytes' page which show histograms. (first page of the analyte report).

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6.9. Standard Uncertainty

The Standard Uncertainty (SU; 'u') statistic has now been added into our reports. The inclusion of this statistic is a requirement for UKAS ISO17043 Accreditation. The SU can be found to the right of the histograms.

The SU is calculated as $1.25 \cdot [SD / \sqrt{n}]$. The target is considered valid if 'u' is less than $0.3 \cdot SD$.

The UK NEQAS Cardiac Markers Scheme Organizer is in agreement with the following statement from Finlay MacKenzie (Birmingham Quality):

"Simplistically, the 'uncertainty' of a target value increases as the number of points used to calculate it decreases. Similarly the 'uncertainty' increases as the SD widens. One would logically have more 'confidence' in a target that was calculated from 100 results with an SD of 2 rather than in a target calculated from only 20 results which had an SD of 10. The SU just puts a number to this common sense notion and so attempts to quantify it in some way. It is reported in the same unit as the mean and SD. It is our reading of the algebra that, when you re-arrange the equations, if $n < 18$ it is impossible to pass."

Reports will reflect this by including this statement:

"Please note that the Standard Uncertainty is not negligible when calculated with < 18 data points." Also reference that a participant should refer to the participant manual.

This statement will appear on the following pages of the report:

- Distribution Summary Page
- Each analytes' page which show histograms. (first page of the analyte report).

6.10. Performance interpretation on reports (Traffic lights and arrow indicators)

There are only three places where the report used colour coding, namely:-

- performance summary icons page
- coloured circle 'traffic lights' associated with the A, B and C scores
- coloured symbols associated with the Specimen % bias

In addition with the traffic light circles there is also a range of "Am I getting better?" trend arrow. The colours have been mapped to the existing familiar grey shadings used on graphs. So, Green equates to the 'white shading', Yellow to the 'light grey shading' and Red equates to the 'dark grey shadings'.

Colour coding uses the universal Green is 'Good', Red is 'Bad' traffic light approach. Yellow is the 'Warning' zone, so though not as 'Good' as it could be, is not yet in the 'Bad' category.

Graphs are in black and white, as it was thought to be it is easier on the eye and so when colour is used it does clearly stand out.

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- **Coloured circle 'traffic lights' associated with the A, B and C scores**

Because the A score is an across-analyte comparator, the limits used for the A score are common across all analytes, namely:

up to and including 100 is white/Green
 from 101 up to and including 200 is light grey/Yellow and
 greater than 200 is dark grey/Red.
 For the B and C scores there are analyte specific settings.

- **Coloured symbols associated with the Specimen % bias**

At the specimen level, we have used a combination approach of colour and symbol together to produce a self evident representation. We have Red double triangles, Yellow single triangles (both of which can point up to represent a positive bias or can point downwards to represent a negative bias) and a Green diamond. The Green diamond indicates the bias of a result which is within desirable limits.

In the following example you can see that the results on Specimen 211A were as follows: very low for Troponin I and BNP was close to the target. Results for specimen 211B were as follows: a bit low for Troponin I and BNP was close to the target. Results for specimen 211C were as follows: a very low for Troponin I and BNP was close to the target.

	Specimen	Pool	Result	Target	Specimen %bias	A score	B score	C score	A	B	C
Cardiac Troponin I (ng/L [pg/mL])	211A	CMS58	138.8	170.41	-18.6	76	-1.1	10.5	● ↘	● ↗	● ↘
	211B	CMS55	154.9	176.04	-12.0						
	211C	CMS57	70.8	91.76	-22.8						
B Type Natriuretic Peptide (ng/L [pg/mL])	211A	BPS23	229.4	247.3	-7.2	110	-5.5	13.0	● ↔	● ↔	● ↔
	211B	BPS22	20.1	17.90	+12.3						
	211C	BPS17	3428.4	3789	-9.5						

- **The "Am I getting better?" trend arrows associated with the A, B and C traffic lights**

Are a broad-brush indicator of whether things are improving (trend arrow up), things are staying the same (trend arrow flat) or things are getting worse (trend arrow down).

There are two things to note. Firstly, we have used the convention 'Trend arrow up' to indicate improvement. So if your B score changes from being +50% to +25% you are improving so the trend arrow is up, even although the B score itself has a lower numerical value.

Secondly, we have used a broad-brush approach not an absolute cut-offs for our trend arrows. So, if your A score has worsened from 125 to 126, we nevertheless categorise this as a 'trend arrow flat' as we do not consider this to be a significant change. If the change does exceed our limits, for example an A score changing from 125 to 145, then this will signify that things are genuinely changing for the worse and a 'trend arrow down' is appropriate.

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7. Performance Criteria - Limits of Acceptable Performance and Performance Problems

7.1. Performance Criteria

Performance criteria have been agreed with the scheme steering committee and NQAAP for Chemical Pathology. They have been implemented by the Scheme Organiser. The following performance limits are reviewed annually – next review date March 2020.

Limits of Acceptable Performance (Lab Based Schemes)

ANALYTE	B Score	C Score
Cardiac Troponin I	±15%	<20%
Cardiac Troponin T	±15%	<20%
Cardiac Troponin 4 th Sample (low Levels)	±15%	<25%
CKMB (Mass)	±20%	<25%
CKMB (Activity)	±25%	<35%
Myoglobin	±20%	<25%
BNP	±15%	<20%
NT-proBNP	±15%	<20%

Limits of Acceptable Performance (POC Based Schemes)

ANALYTE	B Score	C Score
Cardiac Troponin I	±20%	<25%
Cardiac Troponin T	±20%	<25%
CKMB (Mass)	±20%	<25%
Myoglobin	±20%	<25%
BNP	±20%	<25%
NT-proBNP	±20%	<25%

The current limit of acceptable performance for each scheme and analyte may be found at the top righthand area of the 'Histogram' page of the scheme report.

7.2. Definitions of Unacceptable Performance

The definitions of "poor performer" and "persistent poor performer" and procedures when unacceptable performance has been detected have been approved by the NQAAP and our Specialist Advisory Group and will be reviewed annually.

Participants will be defined as **poor performers** under the following circumstances:

- Failure to return for one distribution unless valid reason for non-return has been communicated to the Scheme Organizer
- Having an average **B Score** out- with the stated limits
- Having an average **C Score** out-with the stated limits

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Participants will be defined as **persistent poor performers** under either of the following circumstances:

- A poor performer as defined above, compounds the errors by failing to make more returns or continues with a **B Score** out-with the limits
- **C Score** remains out-with the limits over further distributions.

7.3. Participation and Return Rate

According to NQAAP requirements for acceptable performance, participants are expected to return 100% of results within the relevant cumulative performance scoring period. Where a laboratory is unable to return a set of results, an explanation must be provided.

UK NEQAS Cardiac Markers will assess a laboratory's performance as satisfactory if they have in total up to 2 late and / or amended distributions from the last 12.

7.4. Performance Surveillance and Advisory Panel Liaison

The Scheme Organiser highlights out-of-consensus performance in the routine report by the use of scores, symbols, graphs, banners and 'traffic lights'. The fact of UK NEQAS Cardiac Markers providing participants with a red traffic light constitutes a formal communication from UK NEQAS Cardiac Markers of out of consensus performance. UK NEQAS Cardiac Markers also requests participant complete and return an incident form within a stated timeframe.

It is the responsibility of the Participant to act on, investigate and resolve all out-of-consensus performance. It should be logged in their Quality Management System. For any red B or C score traffic light, participants are expected to both acknowledge and fix the problems as part of their routine Quality Management System.

This is a mutually-beneficial exercise as the laboratory's root-cause analysis provides it (and UK NEQAS for Cardiac Markers) with evidence as to why the error(s) happened and any further action(s) which are required, and provides evidence of compliance with ISO 17043:2010 and ISO 15189:2012. UK NEQAS Cardiac Markers is here to help. If participants are unsure as to why they have out-of-consensus problems or if they are having difficulties with their Root Cause Analysis, then they should contact us.

UK NEQAS Cardiac Markers are required to report to NQAAP for laboratories who's performance is persistently unacceptable. Initially UK NEQAS Cardiac Markers will make contact with the participant inviting them to discuss action to correct the poor performance. If a satisfactory response is made and improvement in performance ensues, no further action is taken. If poor performance persists or no response is made, then a Panel letter (direct from Panel Chairman to Head of Department with lab identity and UK NEQAS lab code disclosed) is written requesting that decisive action is taken to re-establish satisfactory performance; this may include a site visit by Panel members. If this fails, the Joint Working Group may take further action.

Where poor performance is purely method-related (e.g. all users have a large positive or negative bias), the Scheme Organiser may work directly with manufacturers to assist, where possible, with correction of any problem; analogous procedures are in place for apparently IVD-related problems (see JWG guidelines).

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8. Complaints and Appeals Procedure

UK NEQAS Cardiac Markers welcome the opportunity to discuss any problem or query concerning our services

8.1. Complaints Procedure

Should a participant have a complaint regarding any aspect of the scheme, they should notify the Scheme Organiser (alan@ukneqas-cm.org.uk) or Quality Manager (maggie@ukneqas-cm.org.uk). All complaints will be formally responded to within 14 days.

If the matter cannot be resolved at Scheme level, the participant will be advised to refer the complaint to either: a) the Chair of UK NEQAS Steering Group (Clinical Chemistry) b) Chair of the Immunoassay Specialist Advisory Group (i-SAG) or c) National Quality Assurance Advisory Panel. All complaints will be monitored and reviewed by the Immunoassay Specialist Advisory Group (i-SAG). At all times during the complaints procedure, participant confidentiality will be maintained. Participants are requested to assist in this respect.

8.2. Appeals Procedure

An appeal may be raised if participants are unhappy with their performance. Participants can appeal by contacting the Scheme Organiser (alan@ukneqas-cm.org.uk). It is hoped that resolution will occur at this stage however, if participants wish to escalate the appeal, they are referred to the Chairperson of the Immunoassay Specialist Advisory Group (iSAG) of UK NEQAS. Appeals are dealt with in the same manner and timescales as complaints (see above). Appeals are logged in accordance with the Communication Process.

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Appendix 1 - Steering Committee/Specialist Advisory Group – Terms of Reference

To advise the Scheme Organiser on the overall design and operation of the Scheme(s), including aspects such as:

- Appropriateness of the investigations.
- Surveyed nature of the specimens distributed.
- Number and frequency of specimen distribution.
- Source of target values.
- Data analysis and performance assessment.
- Data presentation.
- Communication with participants, including meetings, newsletters, educational activities.
- Communication with the diagnostics industry.
- Research and development for the Scheme(s).
- In consultation with the Scheme Organiser, to liaise with the relevant National Quality Assurance Advisory Panel in setting performance criteria.
- To consider, and advise the Scheme Organiser(s) on, the need for initiation or termination of EQA services for investigations in the area covered.
- To review Schemes' annual reports.
- To receive any representations, to Chairman, members or Organiser, from participants concerning the Schemes.
- To advise the UK NEQAS Board, and where appropriate other relevant organisations (e.g. Department of Health, Joint Working Group on Quality Assurance, UKAS, Medical Devices Agency, Royal College of Pathologists), on any aspect of EQA or quality assurance in the area covered.

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Appendix 2 - Steering Committee – Current Membership

Membership UK NEQAS Steering Committee (Clinical Chemistry)

Member	Position/SAG	Location	Year of Appointment
Prof. I.J. Young	Chairman	Belfast	2010
Jane French	Secretary	Birmingham	-
Dr Neil Anderson		Nuneaton	2012
Dr Bill Bartlett		Dundee	2012
Dr Jim Bonham	<i>Chair Paediatric Investigations SAG</i>	Sheffield	-
Dr Robin Braithwaite	<i>Chair Trace Elements SAC</i>	London	-
David Cameron		Glasgow	2006
Dr Julia Forsyth	<i>Chair General Clinical Chemistry SAG</i>	Derby	-
Mr Ian Hanning		Hull	2012
Dr Anne-Marie Kelly	<i>Chair Interpretative Comments SAG</i>	Manchester	-
Finlay MacKenzie	<i>UK NEQAS Executive/Director Birmingham Quality</i>	Birmingham	
Dr Les Perry	<i>Chair Immunoassay SAG</i>	London	-
Dr Cathie Sturgeon	<i>Director UK NEQAS Edinburgh</i>	Edinburgh	-
Dr Zahra Khatami		Romford	2012
Vacancy	Replacement being sought.		
Dr. Pat Twomey	NQAAP [Observer]	Ipswich	-
Dr. David James	NQAAP [Observer] (Alternate)	Taunton	-

UK NEQAS Scheme Organisers and managers are invited to attend.

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Appendix 3 - Specialist Advisory Groups – Current Membership

Membership UK NEQAS Specialist Advisory Group (Immunoassay) (iSAG)

Name	Position	Location	Year of appointment
Dr L Perry	Chair/Expert Member	Croydon	2007
Dr G Wark	Secretary/Organiser/Expert member	Guildford	-
Dr L Bailey	Expert Member	Liverpool	2006
Professor S Ball	Clinical/Scientific Advisor	London	-
Dr J Barth	Expert Member	Leeds	1990
Professor P O Collinson	Clinical/Scientific Advisor	London	2016
Dr C Evans	Expert Member	Cardiff	2008
Professor W D Fraser	Clinical/Scientific Advisor	Norwich	2011
Ms J French	Co-opted	Birmingham	2015
Dr K Gordon	Expert Member	Basingstoke	-
Dr D Halsall	Expert Member	Cambridge	-
Professor B Keevil	Expert Member	Manchester	2008
Mr F MacKenzie	Organiser/Expert Member	Birmingham	-
Dr R Marrington	Organiser/Expert Member	Birmingham	2016
Dr M Moore	NIBSC Liaison	Potters Bar	2019
Ms L Owen	Expert Member	Manchester	-
Mrs D Patel	Tumour Marker Liaison	Sheffield	2016
Mr A Reid	Organiser/Expert Member	Glasgow	2012
Dr F Riddoch	Expert Member	London	-
Mr M. Roch	Organiser	Birmingham	2018
Professor M Strachan	Clinical Advisor	Edinburgh	2017
DR C Sturgeon	Organiser/Expert Member	Edinburgh	-
Dr C Webster	Expert Member	Birmingham	2018
Dr P Twomey	NQAAP(Observer)	Ipswich	2015

Membership UK NEQAS Cardiac Markers Specialist Advisory Sub-group (cSAG)

Name	Position	Location	Year of appointment
Professor P O Collinson	Chair/Clinical/Scientific Advisor	London	2019
Mr A Reid	Secretary/Organiser/Expert member	Glasgow	2019
Dr L Perry	Expert Member	Croydon	2019
Professor N Mills	Clinical/Scientific Advisor	Edinburgh	2019
Professor R Body	Clinical/Scientific Advisor	Manchester	2019

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Appendix 4 - Joint Working Group for Quality Assurance in Pathology, August 2010

The Joint Working Group for Quality Assurance (JWG) is a multidisciplinary group accountable to the Royal College of Pathologists for the oversight of performance in external quality assurance schemes (EQA) in the UK. Membership consists of the Chairmen of the National Quality Assurance Advisory Panels (NQAAPs), and representatives from the Institute of Biomedical Sciences, the Independent Healthcare Sector, the Department of Health and UKAS.

1. The Head of a laboratory is responsible for registering the laboratory with an appropriate accredited EQA scheme.
2. The laboratory should be registered with available EQA schemes to cover all the tests that the laboratory performs as a clinical service.
3. EQA samples must be treated in exactly the same way as clinical samples. If this is not possible because of the use of non-routine material for the EQA (such as photographs) they should still be given as near to routine treatment as possible.
4. Changes in the test methodology of the laboratory should be notified in writing to the appropriate scheme organizer and should be reflected in the EQA schemes with which the laboratory is registered.
5. Samples, reports and routine correspondence may be addressed to a named deputy, but correspondence from Organizers and NQAAPs concerning persistent poor performance (red – see below) will be sent directly to the Head of the laboratory or, in the case of the independent healthcare sector, the Hospital Executive Director.
6. The EQA code number and name of the laboratory and the assessment of individual laboratory performance are confidential to the participant and will not be released by Scheme Organizers without the written permission of the Head of the laboratory to any third party other than the Chairman and members of the appropriate NQAAP and the Chairman and members of the JWG. The identity of a participant (name of laboratory and Head of Department) and the tests and EQA schemes for which that laboratory is registered (but not details of performance) may also be released by the Scheme Organizer on request to the Health Authority, Hospital Trust/Private Company in which the laboratory is situated after a written request has been received.
7. A NQAAP may, with the written permission of the Head of a laboratory, correspond with the Authority responsible for the laboratory, about deficiencies in staff or equipment which, in the opinion of the NQAAP members, prevent the laboratory from maintaining a satisfactory standard.
8. Laboratories' EQA performance will be graded using a traffic light system; green will indicate no concerns, amber poor performance, red persistent poor performance, with black being reserved for the tiny number of cases that cannot be managed by the Organizer or NQAAP and that have to be referred to the JWG. The criteria for poor performance (amber) and persistent poor performance (red) are proposed by the EQA scheme Steering Committee in consultation with the EQA Provider/Scheme Organizer and approved by the relevant NQAAP.
9. When a laboratory shows poor (amber) performance the Organizer will generally make contact with the participant in accordance with the Scheme Standard Operating Procedure for poor performance. Within 2 weeks of a laboratory being identified as a persistent poor performer (red), the Organizer will notify the

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Chairman of the appropriate NQAAP together with a resume of remedial action taken or proposed. The identity of a persistently poor performing laboratory (red) will be made available to members of the NQAAP and JWG. The NQAAP Chairman should agree in writing any remedial action to be taken and the timescale and responsibility for carrying this out; if appropriate, this letter will be copied to accreditation/regulatory bodies such as CPA (UK) Ltd, UKAS and HFEA who may arrange an urgent visit to the laboratory. Advice is offered to the Head of the Laboratory in writing or, if appropriate, a visit to the Laboratory from a NQAAP member or appropriate agreed expert may be arranged.

10. If persistent poor performance remains unresolved (black), the NQAAP Chairman will submit a report to the Chairman of the JWG giving details of the problem, its causes and the reasons for failure to achieve improvement. The Chairman of the JWG will consider the report and, if appropriate, seek specialist advice from a panel of experts from the appropriate professional bodies to advise him/her on this matter. The Chairman of the JWG will be empowered to arrange a site meeting of this panel of experts with the Head of the Department concerned. If such supportive action fails to resolve the problems and, with the agreement of the panel of experts, the Chairman of the JWG will inform the Chief Executive Officer, or nearest equivalent within the organisation of the Trust or Institution, of the problem, the steps which have been taken to rectify it and, if it has been identified, the cause of the problem. The Chairman of the JWG also has direct access and responsibility to the Professional Standards Unit of the Royal College of Pathologists. Should these measures fail to resolve the issue, the laboratory will be referred to the Care Quality Commission for further action.
11. Problems relating to EQA Schemes, including complaints from participating laboratories, which cannot be resolved by the appropriate Organizer, Steering Committee or NQAAP, will be referred to the Chairman of the JWG.

Joint Working Group for Quality Assessment in Pathology members July 2019.

<https://www.rcpath.org/profession/committees/jwgqa.html>

Mr Nigel Coles - IBMS, alternative representative	Prof Tim Reynolds - Immediate Past Chair, JWGQA
Mr Ben Courtney - UKAS representative	Dr David Ricketts - IBMS representative
Dr Keith Gomez - NQAAP in Haematology, Chair	Dr Lance Sandle - Vice President (Professionalism)
Dr Mark Gompels - NQAAP in Immunology, Chair	Ms Lorraine Turner - UKAS representative
Dr Glenda Horne - NQAAP in Medical Microbiology, Chair	Dr Bryan Woodward - NQAAP in Reproductive Science, Chair
Dr Stephen Lee - MHRA representative (observer)	Dr Pat Twomey - Chemical Pathology representative
Dr Eddy Maher - NQAAP in Genetics, Chair	Dr Judy Wyatt - NQAAP in Cellular Pathology, Chair
Dr Berenice Lopez - NQAAP in Chemical Pathology, Chair	Vacancy - National Screening Committee representative

National Quality Assurance Advisory Panel for Chemical Pathology (NQAAP) members:

Dr Pat Twomey (Chairman)	Dr B Lopez
Mr D Ames	Dr W Simpson
Dr L Ford	Dr R Still
Dr D James	