

Participant's Handbook

UK NEQAS Guildford Peptide Hormones

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1.0 Service Provided and Scheme Background and Aims

Scheme	Analytes
	Insulin and C-Peptide
Peptide Hormones	Gastrin
	IGF-I and IGFBP-3

The UK NEQAS Guildford Peptide Hormones Scheme has been running for over 40 years and assists participants in monitoring the hormones regulating glucose metabolism, growth and gastric function.

The scheme was established in 1975 with the distribution of samples to United Kingdom hospital laboratories measuring insulin and gastrin. This work was initiated and funded by the Supra-Regional Assay Service (SAS) Peptide Hormone Laboratory, which was part of the Department of Clinical Biochemistry and Clinical Nutrition, St. Luke's Hospital, Guildford, Surrey. During the next 10 years participation in the scheme expanded and in 1983 the scheme was extended to include C-peptide. In the 1990s the use of Insulin-like Growth Factor-1 (IGF-I) and Insulin-Like Growth Factor Binding Protein 3 (IGFBP-3) as clinical diagnostic tools were becoming more widespread and the scheme added these analytes to its repertoire.

In 1996 the laboratory moved to new premises within the Royal Surrey Hospital and continues to work from there today, maintaining a close link with the clinical laboratory to facilitate appropriate specimen collections and maintain methodological updates.

In 1998 the scheme joined the UK NEQAS organisation as an associate scheme. By linking with UK NEQAS there is a formal mechanism for external oversight and additional scientific advice to be provided for the scheme. The aims of the Guildford Peptide Hormones Scheme, which are consistent with those of UK NEQAS, are to:

- Provide professionally led and scientifically based schemes with a primarily educational objective
- Provide regular distributions of specimens
- Provide rapid feedback of performance
- Support participants where problems occur and stimulate the overall improvement in performance among all participating laboratories

In order to meet these objectives, lyophilised human-based samples for insulin, C-peptide, IGF-I, gastrin and IGFBP-3 are distributed every six weeks. Reports contain critical information about bias, reproducibility and, with occasional samples, recovery of added analyte. This is achieved with the use of International Reference Preparations (IRPs) and International Standards (IS), if

they are not available, human recombinant materials. Selected distributions may also contain laboratory surveys or interpretative exercises to highlight current laboratory practice.

2.0 Address and Communications

UK NEQAS Guildford Peptide Hormones

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Surrey Research Park

15 Frederick Sanger Road

Guildford, Surrey

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United Kingdom

Telephone: +44(0)1483 571122 ext 3611

Email: rsch.peptideega@nhs.net

Website: www.surreyeqas.org.uk

The website for reporting a result is www.birminghamquality.org.uk

Please quote your laboratory code number in all communications with the Scheme. If no response is received within 5 working days please make contact again as the email communication may have been lost.

The telephone is staffed between 0900 and 1700 Monday to Friday with an answer machine to pick up messages outside these times. Participants will be asked to give their laboratory code number when contacting the centre and will be asked the nature of their enquiry to allow their call to be transferred to the appropriate member of staff. All calls and the actions taken are logged.

A website for the UK NEQAS organisation and which also gives specific information for other UK NEQAS Centres and Schemes, including Guildford Peptide Hormones is at www.ukneqas.org.uk

3.0 Staffing

Dr Gwen Wark	Tel: +44(0)1483 406715
Scheme Director	Email: gwen.wark@nhs.net
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Scheme Manager	Email: godwin.tetteh@nhs.net
Mr Stuart Kerr	Tel: +44(0)1483 571122 ext 3611
Medical Technical Officer 2	Email: stuart.kerr4@nhs.net
Ms Bahareh Jafari	Tel: +44(0)1483 571122 ext 3611
Medical Laboratory Assistant	Email: bahareh.jafari@nhs.net
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EQA Quality Manager	Email: sbainbridge1@nhs.net
Ms Kimberley Pang Medical Laboratory Assistant	Tel: +44(0)1483 571122 ext 3611 Email: kimberley.pang@nhs.net

4.0 External Regulation of our Services

4.1 Accreditation

The scheme is recognised by the UK NEQAS consortium and operates in accordance with the UK NEQAS Code of Practice (see www.ukneqas.org.uk). Accreditation is undertaken by United Kingdom Accreditation Service (UKAS) according to "ISO 17043:2010 Conformity assessment — General requirements for proficiency testing". The schemes are UKAS accredited proficiency testing provider, UKAS No. 7496.

4.2 UK NEQAS Consortium

The scheme has close ties with other UK NEQAS operations though the UK NEQAS Consortium. All UK NEQAS-designated services comply fully with the UK NEQAS Code of Practice.

4.3 Steering Committees & Specialist Advisory Groups

All EQA providers are required to seek advice from and report to Steering Committees and/or Specialist Advisory Groups. The Clinical Chemistry division of UK NEQAS is presently served by an overall Steering Committee (SC) which advises on overall policy matters, with Specialist Advisory Groups (SAGs) providing external scientific advice.

4.4 National Quality Assurance Advisory Panels

All full UK NEQAS schemes report to the National Quality Assurance Advisory Panel (NQAAP) for Chemical Pathology. The names of SC, SAG and Panel Chairs and Secretaries are available on the

UK NEQAS website for any participants who wish to express comments or concerns about schemes and their operation.

5.0 Enrolment and Charges

Prospective participants should contact the scheme by phone or email at the contact details given above for a copy of the current registration form which includes details of the fees for participation.

The scheme's calendar year runs from April to March, although participants are able to register at any time of the year on a pro-rata basis. Participation begins at the first distribution following receipt of completed forms.

Although the majority of participants are diagnostic service laboratories, all laboratories are welcome to join. This includes non-UK, research and in vitro diagnostic medical devices (IVD) manufacturers' laboratories. For UK clinical service laboratories, the act of enrolling in a scheme confirms their willingness to be bound additionally by the current Joint Working Group (JWG) Conditions of Participation. The terms and conditions, and other information about the Joint Working Group can be accessed via the link below:

https://rcpath.org/profession/committees/jwgqa.html

Participation of non-UK laboratories may be subject to the availability of suitable specimen transport. Manufacturers may participate on an 'information only' basis, i.e. without receiving samples and returning results. They may also register methods under development on an anonymous basis.

Between January and March of each year, participants are sent a registration form and requested to confirm or amend their registration details for the following year.

Please inform the scheme immediately if there are any changes to your registration details at any time throughout the scheme year.

If you wish to cancel your participation in the scheme, please notify the scheme in written form. Temporary suspension in the scheme can apply, e.g. if your laboratory is no longer offering the test as a clinical service, provided that the scheme is notified in writing. Failure to provide payment for enrolment in the scheme will result in the cancellation of registration.

The scale of charges is published annually and is available on request. The charging period is 1st April to 31st March, or pro-rata for part year participation. EQA services are run according to the not-for-profit terms of the UK NEQAS Code of Practice. Changes to charges are implemented only after approval by the UK NEQAS Board of Directors.

6.0 Scheme Organisation

6.1 Laboratory code numbers

Participants are assigned a unique laboratory code number, which is common across a number of UK NEQAS schemes. A participant will be assigned an additional laboratory number if more than one result is returned for a single analyte. This may occur if more than one instrument is used for a particular analyte, or if the participant is evaluating a different method in addition to the established method.

6.2 Method codes

Methods are normally referred to by full name, but occasionally a code is used where space is limited on the printed page. Please check your method is up to date and inform us of any changes.

6.3 Confidentiality

The participation, raw data and performance scores are confidential between the individual laboratory and UK NEQAS staff. Performance scores (and some relevant raw data) may be shared with the relevant Advisory Panel under defined circumstances (see section 5.0 Enrolment and Charges) as part of the routine reporting of persistent poor performance. These data may be shared with local management, regional QA officers, accrediting bodies and suppliers of equipment and reagents where appropriate and necessary, but only with the participant's permission. Reports are copyright and may not be copied, distributed, published or used for publicity and promotion in any form without written consent of the Scheme Director on each and every occasion. However, performance data may be shared with individual client's e.g. GP's without consultation with the Scheme Director.

7.0 Scheme Operation

7.1 Specimens

Specimens are obtained from three sources;

- Endogenous serum either from donations, purchased serum, or Pooled patient samples.
- Serum spiked with the analyte(s) (Insulin, C-peptide or IGF-1) to clinically relevant concentrations.
- Serum spiked with peptides or chemicals known to interfere with methods for the analytes to ensure participants are aware of possible interferences on their method.

Specimens may be "spiked" with standards or other source of analyte, or with another analyte(s) to test method performance. 1 mL aliquots of the sample pools are lyophilised and stored at

4°C prior to issue. No preservative is added to the lyophilised pools. Specimens are distributed at ambient temperature. Specimens must be reconstituted with 1 mL of distilled or deionised water and mixed for 15 minutes before analysis by participants.

Several analytes share a common sample. The sample pools are prepared with:

- Insulin and C-Peptide
- Gastrin
- IGF-I and IGFBP-3

Therefore a laboratory that is registered for insulin, C-peptide and IGF-I will receive 2 sample sets.

The specimens are provided solely for the purposes of EQA. Residual material may be retained by participants for method evaluations. However it is recommended that fresh samples are obtained from the scheme for such evaluations.

If specimens are to be used in research which is expected to be published, written consent must be obtained from the Scheme Director on each and every occasion. The table below shows the anticipated range of analyte concentrations that can be used by the scheme:

Programme 1						
Insulin	<10 - 1000	pmol/L				
C-peptide	<100 - 4000	pmol/L				
Programme 2						
Gastrin	20 - 1000	mU/L				
	Programme 3					
IGF-1	1 - 70	nmol/L				
IGF-BP3	2 - 7	mg/L				

Safety precautions in handling specimens

The clinical materials used to produce the EQA samples are screened for COVID-19 prior to sample production. However, as for all clinical material, EQA samples should be handled as if being capable of transmitting infections. The same health and safety precautions which are normally adopted in the handling of patient specimens should be used during EQA sample receipt, storage, preparation for analysis, and their eventual disposal.

7.2 Schedule of specimen and report distribution

Specimens are distributed every 6 weeks together with a form for reporting results to the registered scheme contact. In the UK, first class mail is used. For overseas participants, packages are posted by airmail with an express surcharge (if necessary). A schedule of specimen despatch dates is provided each year and available on the UK NEQAS Birmingham Quality website (www.birminghamquality.org.uk). Participants should contact the scheme at the contact details given in section 2.0 if specimens are not received within 7 working days of the distribution despatch date.

Sample packaging complies with current UN3373 requirements for the postage of clinical material.

Interim reports from the previous distribution are sent out following the closing date for the distribution. These reports should be received by participants within 5 working days of the result return deadline. The final reports are made available approximately 6 weeks later with the interim reports of the following distribution. Please contact the scheme if reports have not been received within this time period. The scheme is also due to introduce Network Reports in early 2021. These reports will allow users with multiple participant numbers (multiple analysers performing the same tests) to easily compare instrument performance on one combined report as well as allowing access to the normal instrument specific reports from a single centralised login. Please enquire by email if this is a service you would be interested in utilising.

8.0 Processing UK NEQAS Specimens

8.1 Receipt and analysis

Please contact us immediately if you receive incorrect or damaged specimens and replacements will be sent.

It is recommended that if an assay is not to be performed on the day of receipt the lyophilised specimens should be stored at 4°C.

Specimens must be reconstituted with 1mL of distilled or deionised water and mixed for 15 minutes before analysis.

If reconstituted samples are required for further analysis, the samples should be stored frozen. UK NEQAS samples are intended to monitor your performance for routine patient specimens, so please process them through your normal reception, analytical and reporting procedures. In order to meet this goal, the following guidelines should be met:

- EQA samples should be labelled in the same manner as a patient sample it should not be known by the analysts to be an EQA sample.
- Instrument calibration, routine maintenance, and other assay parameters should be done
 with no greater frequency for EQA samples than for patient samples.
- EQA samples should be run the same number of times as a patient sample it should not be run multiple times UNLESS a patient sample is run multiple times under the same circumstances. If it is desired to run replicate determinations of the EQA sample, the results should be returned BEFORE the additional replicate assays are performed.
- Results below the minimum detection level should be reported to show the limit of detection.

By adhering to these guidelines, the efficacy of external quality assessment is maximised.

8.2 Return of results

Results must be submitted by the closing date on the return form.

In the first instance results should be returned using the password-protected website facility: www.birminghamquality.org.uk. Following enrolment with the scheme, participants will be issued with a username, password and instructions to access the website. Where access to the website is not possible results may be returned by post or email to the Scheme's email address (rsch.peptideeqa@nhs.net). Please ensure results and decimal points are written clearly on the return sheet as there can be a loss of clarity when scanned. Please indicate clearly on the return sheet if your units differ from those shown.

9.0 Performance Assessment

9.1 Failure to return results

If you do not return any results for a distribution by the due date you will still receive a report. However, regular participation is essential if appropriate method performance data is to be obtained and is part of the criteria for good performance.

If results are not returned for a distribution, you will be regarded as having poor performance and it will be noted on your report when issued.

If you are unable to return results on a distribution please contact the scheme as soon as possible with an explanation of the reason for not doing so.

9.2 Target values

UK NEQAS Guildford Peptide Hormones attaches great importance to validation of target values, rather than simply accepting consensus means as the "correct" result. Target values should be accurate and stable, but this is difficult to achieve for peptide hormones, where reference methods are largely unavailable. UK NEQAS Guildford Peptide Hormones aims to meet minimum validity criteria by testing recovery, linearity and stability of the targets at regular intervals throughout the year. Wherever possible the all-laboratory trimmed mean (ALTM) is used as the target. Along side the target value we also provide the standard deviation and the coefficient of variation. Where there are <5 participants that have submitted results the data set is not trimmed and the standard deviation and coefficient of variation are not calculated, this ensures a robust SD while still allowing it to be displayed for smaller method groups.

9.3 Introduction to analytical performance scores

'ABC of EQA' is an ISO 17043:2010 compliant framework for the assessment of a laboratories analytical performance in a particular assay which meets and surpasses the utility of existing systems. The main benefit for participants, EQA Organisers, Steering Committees, Specialist Advisory Groups and the NQA Advisory Panels alike, is that it is a single system, which can allow meaningful comparisons to be made between analytes, schemes and disciplines.

The reports for the UK NEQAS Guildford Peptide Hormone' scheme are structured to best utilise the 'ABC of EQA' scoring system, so you are able to see at a glance if your laboratory is performing well. If performance is acceptable, no further action is required. If performance is poor, you can probe further into the data presented. Similarly, you can see if you are performing in keeping with other users of your method and whether the method itself is performing well.

Definitions

There are three scores A, B and C

A is for Accuracy (total error)

B is for Bias

C is for Consistency of bias

They are conveniently referred to as the 'A score', 'B score' and 'C score', or simply A, B and C. Every laboratory in the scheme will have an A, B and C score for each analyte they measure and all 3 should be used when reviewing performance. Each of the 3 scores is calculated over a rolling time-window and thus comprises results from many specimens. They are always being updated with fresh current data, while older data drops out of the 'time-window'. The time-window has been set at 6 distributions. One of the main purposes of a performance score derived from many samples is to 'smooth out' the natural variation in deviations from target values over a number of distributions, by trimming extreme values and deriving a robust estimate of the central tendency for overall bias together with an index of its consistency. Thus when interpreting the performance score elements of reports, it is important to note that a small number of atypical results are unlikely to affect overall scores, and aberrant results which are numerous enough to affect performance scores will take time to work their way out of the scoring 'window'.

For all UK NEQAS centres, a low score is 'good', a high score is 'bad'.

The A Score Accuracy

The A score is weighted as part of a transformation process to take into account factors such as

'degree of difficulty' and normalised (median set at 100). The A score is primarily used as a quick

'comparator' or 'screening tool' for performance across all analytes. An A score of '100 is

'average', but this may of course be 'better' or 'worse' than what is required clinically, depending

on the analyte. As more UK NEQAS schemes adopt the 'ABC of EQA' approach, the more useful

the A score becomes in allowing broad comparisons to be made between analytes.

The A score tells you, on average, how good your overall performance is. This takes into account

such factors as bias, consistency of bias, degree of difficulty etc. It has been transformed to

ensure that A scores are broadly equivalent across analytes. For example, if you have an A score

of 85 for Insulin and you also have an A score of 85 for Gastrin, this would indicate that you are

performing both, on average, equally well.

The A score is an estimate of accuracy [total error] and is derived by taking the Specimen % bias

and transforming it by a 'degree of difficulty' factor (see below) to get a Specimen transformed

bias [this can be positive or negative]. The modulus of this Specimen transformed bias is then

taken to give the Specimen Accuracy Index [as it is a modulus it has no sign]. Finally, the 'A score'

is calculated as the trimmed mean of all of the Specimen Accuracy Indices in the rolling time-

window.

Because the A score is an across-analyte comparator, the limits used for the A score are common

across all analytes, namely:

Up to and including 100 (green on report)

From 101 up to and including 200 (yellow on report)

Greater than 200 is (red on report)

Degree of difficulty factor: The transformation itself has been empirically derived separately for

each analyte and is based on modelling of data dependent on the concentration (target value)

for the individual specimen. An examination of the relationship between CV and target value for

the analyte was conducted to derive an equation for this relationship. This yielded the

concentration-dependent factors used. Normalisation of the factors to yield a median (average

participant) A score of 100 was then carried out.

The B and C Scores

Bias and Consistency

The B and C scores (which have not been transformed) should be looked at together and provide analytical data on average bias and its consistency (pattern). The B score is Bias and therefore shows, on average (across the 6 distribution window), how far from the target results are and if results are running high or low.

The Consistency of bias or C score indicates, on average, if you usually have the same bias pattern. It is also not transformed and can assist in answering the following questions. 'Do you have different bias depending on the concentration of analyte in the sample?' 'Does your bias vary depending on the specimen matrix?' 'Has your bias changed during the time window?' 'Are you imprecise?'

A high (poor) C score does not necessarily mean that you are imprecise, though if you are imprecise, it is impossible for you to have a very good (low) C score. Poor consistency of bias is not the same as imprecision. The 'C score' is simply the standard deviation (adjusted to take into account the degree of trimming) of the data which make up the B score.

9.4 Calculation of analytical performance scores

A Score:

When calculating the A score, the Specimen Bias for each sample in the 6 distribution window is calculated. The equation for this is given below:

Specimen Bias (%) =
$$\frac{Result-Target}{Target} \times 100$$

The Specimen Biases are the each divided by the appropriate Degree of Difficulty:

Specimen Transformed Bias =
$$\frac{Specimen Bias (\%)}{Degree of Difficulty}$$

Using this, the Specimen Accuracy Index is found.

 $Specimen\ Accuracy\ Index = Modulus\ of\ Specimen\ Transformed\ Bias$

The trimmed mean of the specimen accuracy indices gives the A score.

B Score

The B score is the average (trimmed mean) Specimen Bias across the 6 distribution window. The equation for the Specimen Bias is given below:

Specimen Bias (%) =
$$\frac{Result-Target}{Target} \times 100$$

C Score

The C score is the standard deviation of the biases cross the 6 distribution window.

The Scheme Director / Manager may exclude certain sample pools and/or methods from the calculation of target values and scoring if these are atypical or may unduly affect apparent assay performance. These actions may be performed in consultation with the Specialist Advisory Group (SAG) for Endocrinology and Immunoassay.

The Standard Uncertainty

The Standard Uncertainty (SU) statistic has now been added into our reports. The inclusion of this statistic is a requirement for UKAS ISO17043:2010 Accreditation. The SU can be found to the right of the histograms.

The SU is calculated as outlined in ISO 13528 using the formula 1.25*[SD/SQRT n]. The 'n' used in this calculation relates to a post trimming value (where appropriate) not the 'n' value listed on reports. The target is considered valid if 'u' is less than 0.3*SD. It is our reading of the algebra that, when you re-arrange the equations, if n<18 it is impossible to pass. As such any results with a n<18 should be interpreted with caution.

9.5 Late results

We will accept late returns to the interim reports but once the final reports are published, late results can only be accepted in limited situations. All amendments to interim or final reports are made at the discretion of the Scheme Director / Manager.

9.6 Blunders and their correction

Blunders are defined as errors, which may or may not be detected as outliers, and a record is kept by the scheme of each incident. Participants are allowed one blunder per scheme year, depending on the circumstances and risk, they may be considered poor performance. Blunders may be due to:

- assaying the wrong samples
- assaying the right samples in the wrong order
- incorrectly transcribing laboratory results from computer systems or worksheets to the results document
- use of incorrect units and/or conversion factors
- technical errors e.g. incomplete mixing after thawing, faulty sampling/pipetting, incorrect preparation of calibration solutions etc.

Such errors will be corrected in most circumstances, so that they do not confuse the underlying assay performance. However, the fact that blunders have occurred will be recorded separately. Amended reports will be given a unique identification with reference to the original report and include a statement concerning the reason for the amendment.

9.6.1 Amendment prior to reporting deadline

For Amendments prior to the reporting deadline, amended copies of already entered results should be clearly marked "Amended results" with the change unambiguously highlighted and returned to us by email.

9.6.2 Amendments after the reporting deadline

Please email us to explain the problem. Results can be amended prior to the publication of the interim reports.

Once interim reports have been published, amendments should be requested in writing (by email) with an explanation for the reason for any amendment. All amendments are made at the discretion of the Scheme Manager or Director.

Where investigation reveals the cause of the error and repeat results are available, correction of the original results is permissible. A copy or screenshot of the experiment with evidence of the results and analysis date will be requested. However, the fact that incorrect results were reported will be recorded as a blunder.

Once a final report has been issued, amendments to results will not be accepted useless exceptional circumstances can be demonstrated. As before, these amendments are made at the discretion of the Scheme Manager or Director.

9.6.3 Amendments after the receipt of reports

These should be reported in writing with an explanation of the reason for any amendment. Please include a description of how patient samples are run in your laboratory giving particular attention to the area that caused the blunder, e.g. how your results are reported if the blunder was caused by a transcription error. Changes can be made only in those cases where the error is an artefact of running the EQA samples differently to patient samples. If at all possible, EQA samples should be processed exactly the same way as patient samples from labelling to sending the results back on the normal laboratory results forms. Errors caused by sample mislabelling

cannot be corrected because a similar error could be made on patient samples. Where investigation reveals the cause of the error, and repeat results are available, correction of the original results may be permissible. However the fact that you reported incorrect results will be recorded and reviewed annually.

9.7 UK NEQAS Guildford Peptide Hormones errors

If you suspect that we have made an error please let us know immediately. We audit all such errors and it is important that we know about them so that we can improve our service. Errors made by UK NEQAS Guildford Peptide Hormones will be corrected and amended reports will be provided. Any penalties incurred by the laboratory will be removed.

10. Performance Criteria

Limits for acceptable performance are recommended by the Scheme Director and endorsed by the National Quality Assurance Advisory Panel (NQAAP), after consultation with the schemes and Specialist Advisory Group (SAG), to reflect clinical requirements, the state of the art analysis and the need for regular Quality Assurance monitoring. The criteria include acceptable limits for the B and C scores, and for return rate. These are summarised in Appendix A Limits of Acceptable Performance.

The 6-weekly reports include figures to show your performance in relation to these criteria. Laboratories should aim to maintain performance within these limits and are invited to contact the Scheme if problems appear to be developing, whether in analytical performance or in their ability to maintain return rates.

10.1 Persistent poor performance and action taken

A laboratory is considered to be a persistent poor performer for a given analyte if:

 their B and/ or C scores are outside the performance criteria for three consecutive distributions

Or if

• it fails to return results for three distributions in 8 month period, without notifying the UK NEQAS Centre of a change in participation.

UK NEQAS Guildford Peptide Hormones is required to report to the NQAAP for Chemical Pathology any laboratory performance that is persistently unacceptable.

The Scheme Director will make informal contact with any participant falling into the above categories inviting them to discuss action to correct the poor performance. If a satisfactory response is received and improvement in performance ensues, no further action will be taken. If no response is received or performance fails to improve then the Director will notify the Chairman of the appropriate NQAAP. Advice is then offered to the Head of the laboratory in writing or, where appropriate and very rarely, following a visit to the laboratory from a NQAAP member, or appropriate expert (if agreed).

If poor performance is due to the method used, such that all method users have a large negative or positive bias, the Scheme Director will contact the assay manufacturer and work directly with them to solve the issues with the assay.

11. Past UK NEQAS Specimens

We can usually provide aliquots of previously issued specimens with target values for laboratories wishing to check existing assays or to evaluate new ones. An additional charge may be made for these specimens.

12. Scheme Development & Scientific Support

External scientific advice is provided by the Specialist Advisory Group (SAG) for Endocrinology and Immunoassay. The SAG reports to the UK NEQAS Steering Committee for Clinical Chemistry which co-ordinates UK NEQAS policy and provides strategic direction.

The scheme reports to the National Quality Assurance Advisory Panel (NQAAP) for Chemical Pathology. We are required to report those participants whose performance scores are outside the set limits on a number of occasions or who fail to return sufficient results. The details of the SAG and Steering Committee are available from the UK NEQAS Office while the list of NQAAP panel members is available on the RCPath website via the link below:

https://rcpath.org/profession/committees/jwgqa.html.

13. Comments, Complaints and Appeals

If you have any comments or complaints about any aspect of the scheme, whether scientific or operational, or wish to appeal against assessment of performance, please contact the scheme. The Scheme Director and/or Scheme Manager will follow up on the initial response with a thorough investigation and is ultimately responsible for ensuring that appropriate corrective action has been taken.

Complaints should be in the form of letter or email. The on-line results document includes a section in which participants may include comments or remarks for the attention of the organisers. While these will generally refer to the samples or to analytical difficulties experienced by the participant any observation, at any time, is welcomed. Please use your laboratory code number in all correspondence and provide details of the distribution date and specimen numbers wherever possible.

A formal complaints procedure is in place and wherever possible will be actioned internally. We will endeavour to rectify problems as soon as possible. If the problem cannot be resolved it will be referred to the Chair of the Joint Working Group on Quality Assurance.

Participants may prefer to address comments or complaints, including an appeal against assessment of performance, to any member of the UK NEQAS Specialist Advisory Group for Endocrinology and Immunoassay, the Steering Committee for Clinical Chemistry or the National Quality Assurance Advisory Panel for Chemical Pathology.

14. Subcontracted Services

Various aspects of the proficiency test scheme can from time to time be sub-contracted. When sub-contracting occurs it is placed with a competent sub-contractor and the proficiency testing provider is responsible for this work.

15. Changes to Scheme Design

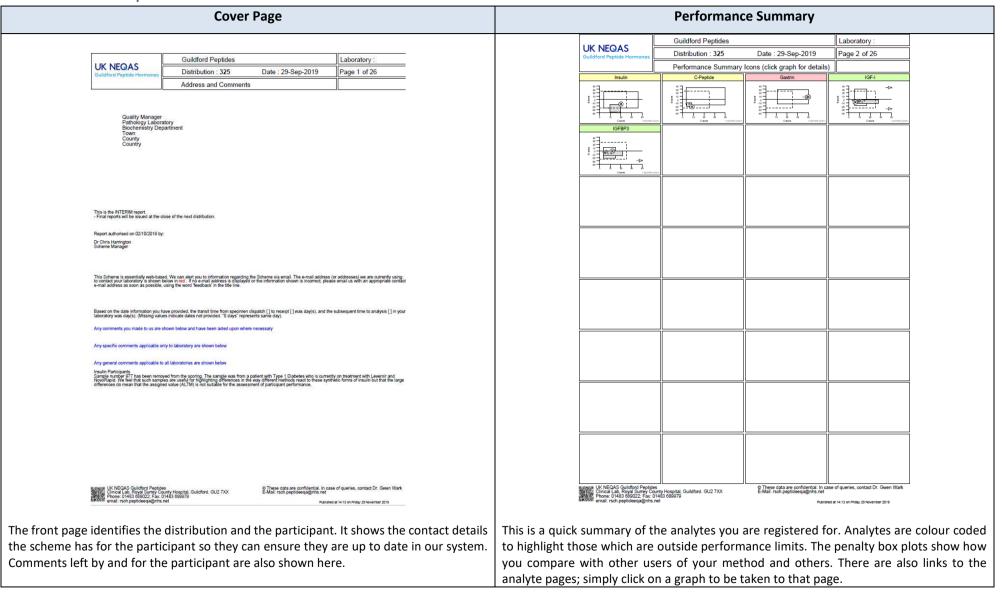
In the event that changes are made to the Scheme, participants will be informed promptly either in writing as a letter posted with the next distribution of samples or by email to the identified contact personnel given on the participant's registration form.

16. Terminology

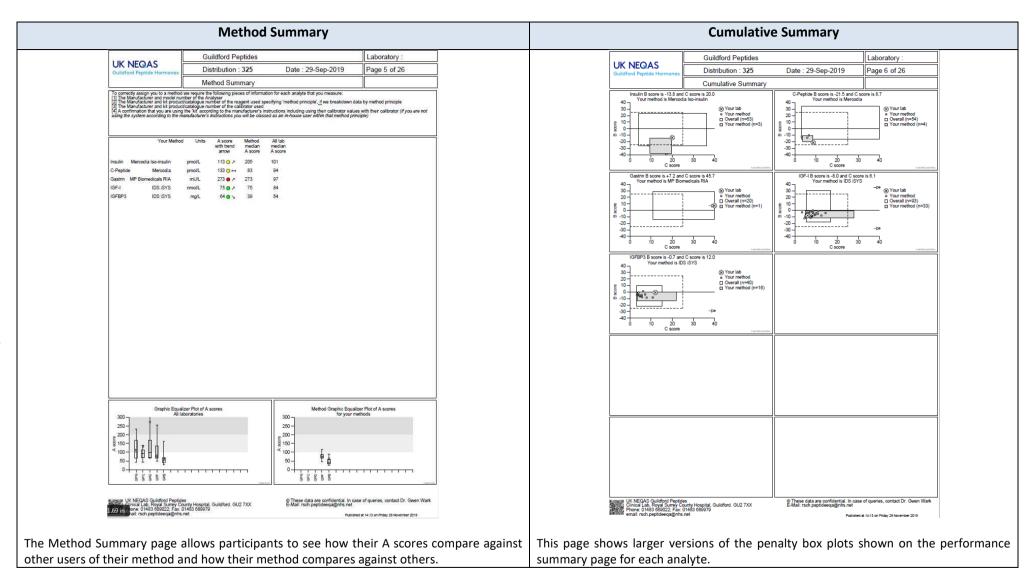
ALTM	The All Laboratory Trimmed Mean, which is the geometric mean of the entire set of trimmed results for a specimen.
Bias	The difference between your result and the target result expressed as a percentage of the target.
Distribution	A group of specimens in a particular scheme that are sent together to each participating laboratory.
Pool	A bulk preparation of serum usually prepared from several individual donations and of defined characteristics.
Sample/ Specimen	An aliquot of a given pool . The same pool may be issued on two or more occasions as different specimens.
SAG	Specialist Advisory Group.
Scoring	The 3 values per analyte which provide an overview of your performance. The scores are the A, B and C scores. More information regarding how they are calculated and how they should be interpreted is available in section 9.
Target Value	The scheme uses the ALTM (see above) to establish the target value.
Transformation	The A score is the only transformed score used to assess participant performance. This is done by dividing the Specimen Bias by the appropriate value for the Degree of Difficulty. This means that this score takes into account how difficult the assay is to perform and how difficult it is at that particular sample concentration before the score is calculated.
Trimming	The effect of aberrant results that may be present is minimised prior to statistical analysis. The chosen method is that of 10% Healy, which involves trimming the lowest and highest 10% of results on datasets where n>4. Note that trimmed results are not necessarily outliers.

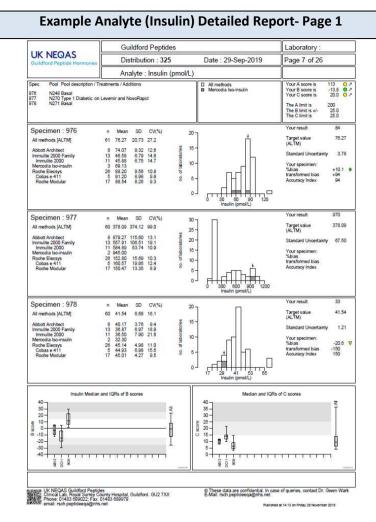
Unusable Specimen	Unusable specimens include those with analyte concentration near the detection limits of the assay, those with added interfering substances or those deemed to not be homogenous. In some circumstances, unusable specimens may be issued by the scheme but will be removed from the scoring and therefore will not be used to assess performance. These samples are for education purposes only and are used to highlight methodological differences.
Usable Specimen	A specimen that has no unusual or unacceptable features will be deemed to be usable for the calculation of ABC scores.

17. Distribution Reports



Participation Summary							Dis	tribu	ıtion	Sum	mary	/							
and the second of the second of	Guildford Peptides Laboratory :							Guildford Peptides								ory :			
UK NEQAS	Distribution : 325	Date : 29-Sep-2019	Page 3 of 26			UK NEQAS Guildford Peptide Hormones		UK NEQAS Guildford Peptide Hormo		Distribution : 325 Date : 29-Sep-2019							Page 4 of 26		
	Participation summary									tion Sum						_			
All our time periods are 'rol You may wish to keep you	e over the last 8 months (rolling time windov ng' to give you current information. wn log of Callendar Year or Financial Year time points sensus performance will be highlighted in red and can performance for: Gastrin	if you require 'year-end' statements for your on be clicked for further details.	vn internal use.	•		If your laborate symbol. It is the action. Please before logging performing as	ory is outside of the responsibility of t do not wait to rece the non-conformit well as the state-of	acceptable he laborator live a formal y and, where the-art allow	limits of pert y to undertal notification of necessary, ws; it does no	formance for ke an internal of performand acting upon to not necessarily	any its rolling to I investigation to be from the Sch the data contain y mean that you	me-window so o establish the neme Organise ned in your rep ur assay / labo	cores (A, B or e underlying o er or the Natio port. A green to pratory perform	C scores), this ause and put in nal Quality Ass raffic light men nance is good o	will be indica place correc urance Advis ly reflects the mough clinical	ted by a red traffic live and preventive ory Panel (NQAAP at your laboratory is ally.	ght		
You have in consensus pe You have no performance		IGF-I IGFBP3																	
Participation and Retu						Insulin (pmol/L)	976 977 978	Pool N248 N270 N271	84 970 33	76.27 378.09 41.54	Specimen %bias +10.1 ♦	A score	-13.8	C score	A B	0.7			
Participation 6 d Late Returns 0 d		Rating Affected Distribution Satisfactory Satisfactory Satisfactory	ns			C-Peptide (pmol/L)	978 976 977 978			41.54 256.5 621.3 375.4	-20.6 ▼ -28.7 ▼ -3.4 ◆ -9.7 ◆	133	-21.5	8.7	0↔ 0+	• • •			
Analytical Performance	e for specimens from distribution 325 only n with your IQC and other QA measures, if your curren					Gastrin (mU/L)	976 977 978	G178 G177 G176	87 53 27	70.5 44.5 17.6	+23.4 △ +19.0 △ +53.0 ≜	273	÷7.2	45.7	07 00	• • •			
Out of consensus for at let in consensus for all specir You have no specimen da	ens for: Insulin	Gastrin IGF-I	GFBP3			(nmol/L)	976 977 978	F217 F215 F216	13.1 13.7 21.5	13.70 14.50 22.28	-4.4 • -5.5 • -3.5 •	75	-8.0	6.1	0/ 0+	○ ↔			
You have no specimen da You are not registered for:						IGFBP3 (mg/L)	976 977 978	F217 F215 F218	4.1 4.2 4.8	3.580 3.710 4.385	+14.5 • +13.2 • +9.5 •	64	-0.7	12.0	0 v 0/	0.0			
	nary page is an exciting new feature which we hope you lytes as well as the existing numerical analytes listed helpful if you could check the accuracy of the data pre		will ultimately expand into				857558	Section 1	.000	::EEFEE	25(2:35.0								
																	_		
emesse UK NEQAS Guilafo G. Cinicat Lab Royal and P. Phone CH483 Sept. a World email: risch peptide	Peptides jurrey Courty Hospital, Guildford, GU2 7XX 2; Fax: 01483 696979 jag@nhs.net	© These data are confidential. In case E-Mail: risch peptideeqa@nins net Published at	of queries, contact Dr. Gwen Wark 14:13 on Friday 25 November 2019			UK NEC Clinical Phone: email: n	QAS Guildford Pep Lab, Royal Surrey 01483 689022; Fa sch.peptideeqa@n	tides County Hos x: 01483 68i hs.net	pital, Guildfo 9979	ord. GU2 7XX	t)	© These di E-Mail; rsci	lata are confid h.peptideeqa(ntact Dr. Gwen Wai	-		
This is an overview of yo	ur average and curre	ent performance	for each anal	yte in addition	This is a sum	mary c	of this d	listril	butio	n. It	incluc	les all	l resu	lts ret	urne	d for th	e distribution,		
to a record of how many	to a record of how many distributions you have returned results for, how often your results			the target r	esults,	pool	and	sam	ple i	denti	fiers,	spec	imen	bias	es and	your current			
were late and how many	amendments you ha	ave required over	the last 8 mc	onths.	performance	score	es. The	traf	fic li	ght s	systen	n ma	kes i	t easy	to :	see wh	ether you are		
			performing a	ccepta	ably or	not.													





This is the first Analyte page for insulin. This is replicated for each analyte the participant is registered for. Here the user can view more detailed breakdowns of data from different methods, histograms showing the distribution of results as well as more information such as the SD of results and the uncertainty of measurement. This report shows an educational sample (977). This is for a patient on treatment for type 1 Diabetes. The sample was issued to ensure that participants understand how their instrumentation is affected by clinical therapy. It was removed from the cumulative performance scoring.

Example Analyte (Insulin) Detailed Report- Page 2

Luc No		Guildford Pe	eptides						Labora	tory:				
UK NE	QAS Peptide Hormones	Distribution: 325 Date: 29-Sep-2019								Page 8 of 26				
ASSESSED TO SECURITY.	24 State of Cong. 100 110 No. 550	Analyte : Ins	ulin (pmol/L)					T						
Pool (exclusion) [Type]	Distribution 320 17-Feb-2019 result target %bias	Distribution 321 31-Mar-2019 result target %bias	Distribution 323 19-May-2019 result target %		Distribution 07-Jul-20 suit target	19	18	ributio -Aug-2 targe		29-Sep-2019				
N275 N274				-8			30 32	37.05 38.73	-19.0 -17.4					
N278 N271 N286 N248 N277 N276 N272 N273 (N270)	29 41.28 -29.7 37 42.19 -12.3 73 73.42 -0.6	23 40.57 -43.3 92 101.16 -9.1 162 160.69 +0.8	89 94.43	-5.8 +5.1	22 41.07 96 213.80 96 375.73	-48.4 -8.3 +5.4	31	41.14	-24.6	33 84 (970)	41.54 76.27 378.09	-20.6 +10.1		
Method mean A score B score	Mercodia Iso-insulin -14.2 161 -24.5 27.8	Mercodia Iso-insulin 173 -17.2 25.2 29.4	Mercodia Iso-insuli 170 -24.7 27.6	13.0	rcodia Iso-in 68 1.9	nsulin -16.4	Mercod 157 -21.6 28.4	ia Iso-i	nsulin -20.3	113 -13.8	dia Iso-in	sulin -5.		
C score		QRs of A score	27.6	40 ¬	9.1	nsulin Me		IIQRs	of B score	20.0				
250 - 200 -	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	111(111	Ī	30 - 20 - 10 - 80 0 - 0 -10 - -20 - -30 - -40 -				1 1	, , ,		Ī			
40 - 10 - 20	fin B score is -13.8 and C /our method is Mercodia	So-insulin Solver la Your la Your n	ethod (n=63) ethod (n=3)	40 35 30 25 20 20 15	• I	Media	n and IQI	Rs of C	Scores		IV I			

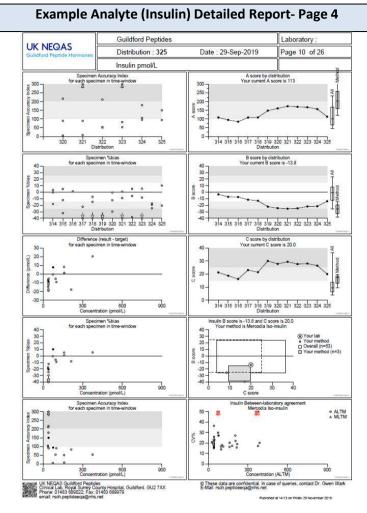
This is the second Analyte page for insulin. This is replicated for each analyte the participant is registered for. This page shows the participant results, targets, and biases of all the samples which make up the ABC scores. It also shows several graphs useful in troubleshooting an assay.

Example Analyte (Insulin) Detailed Report- Page 3 Guildford Peptides Laboratory **UK NEQAS** Page 9 of 26 Distribution: 325 Date: 29-Sep-2019 Analyte: Insulin (pmol/L) Mean SD CV(%) Mean SD CV(%) Mean SD CV(%) 76.27 20.73 27.2 378.09 374.12 99.0 41.54 879 27 115 60 13 1 Beckman Access DiaSorin Liaison Immulite 2000 Family 67.45 72.70 46.58 45.88 69.90 65.00 69.13 85.00 88.20 76.14 91.20 88.54 81.36 557.91 106.51 19.1 584.89 63.74 10.9 ortho Vitros Roche Elecsys 1010 + 2010 Cobas e 411 Roche Modular Siemens Centaur 6.98 15.5 4.27 9.5 -0.6 -12.3 9.6 6.4 -3.2 -16.1 -4.4 -14.4 -7.5 -25.6 +6.6 +11.8 +8.4 +19.1 +11.6 -5.6 -9.7 -19.8 -4.7 -19.1 -16.6 -24.2 -9.4 -32.5 +6.6 +6.5 +8.4 +12.1 +6.9 -6.4 79 125 45 140 130 210 76 205 59 101 79 159 101 +4.4 -12.2 -4.1 -3.8 -20.6 -5.5 -19.7 +6.6 +22.6 +8.4 +23.7 +22.6 -4.4 10.9 5.3 5.8 13.9 13.4 14.3 9.0 17.2 7.3 6.4 7.7 9.1 6.1 24.2 89 170 47 182 164 223 85 258 59 183 79 192 183 155 11.3 6.2 5.9 14.8 15.8 14.3 9.9 18.6 7.3 9.2 7.7 9.6 7.0 37.4

This is the third Analyte page for insulin. This is replicated for each analyte the participant is registered for. It further breaks down the different methods and provides you with detailed information about how the methods compare. This is particularly useful when troubleshooting assays or when looking to purchase new equipment.

© These data are confidential. In case of queries, contact Dr. Gwen Wark E-Mail: rsch peptideega@nhs.net

RUDESID UK NEQAS Guildford Peptides Guildford Surrey County Hospital, Guildford, GU2 7XX 27.25 Phone: 01483 689022; Fax: 01483 689979



This is the fourth Analyte page for insulin. This is replicated for each analyte the participant is registered for. The final page shows a number of graphs including how scores have changed over time and how your lab and method compare against other participants. This is particularly useful when troubleshooting assays.

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Appendix A: Limits of Acceptable Performance

A Score	
<100	Better than average performance (Green)
101 - 200	Worse than average performance but within acceptable performance limits (Amber)
>200	Worse than average performance and outside acceptable performance limits (Red)

B Score	
<±25%	Acceptable performance
>±25%	Unacceptable performance

C Score	
<25%	Acceptable performance
>25%	Unacceptable performance

Participants will be defined as poor performers under the following circumstances:

- Having an average B Score out- with the stated limits
- Having an average C Score out-with the stated limits
- Failure to return for 1 or more distributions in a 6 distribution period unless valid reason for nonreturn has been communicated to the Scheme.
- Returning results late for 2 or more distributions in a 6 distribution period
- Having 2 or more blunders in a 6 distribution period